

# Influence of phosphate ion on the fluorescence of 3-fluorotyrosine

Short Communication

S. I. Vdovenko, M. T. Kolycheva, I. I. Gerus, and V. P. Kukhar

Institute of Bioorganic and Oil Chemistry, Academy of Sciences of Ukraine, Kiev, Ukraine
Accepted August 8, 1992

Summary. 3-Fluorotyrosine fluorescence is quenched effectively by phosphate ions not only by a dynamic but also by a static mechanism owing to H-bond complex formation in ground state. 3-Fluorotyrosine  $pK_a$  values both in the ground and first excited state (8.3 and 4, respectively) are appreciably lower than those of tyrosine, thus promoting 3-fluorotyrosinate ion formation in the excited state. Additional emission owing to 3-fluorotyrosinate ion (near 350 nm) may be taken erroneously for tryptophan fluorescence.

**Keywords:** Amino acids – 3-Fluorotyrosine – Fluorescence – Quenching – Absorption – Phosphate

## Introduction

The introduction of fluorinated aromatic amino acids into membrane proteins is regarded as extremely effective method of modification (Ghosh et al., 1985; Kuriatov et al., 1984). In particular, the substitution of native tyrosine residues by the fluorinated ones leads to little disturbances of peptide conformation (Kuriatov et al., 1984). In some cases fluorine substitution of aromatic amino acids shifts the fluorescence spectra and increases quantum yields (Chen et al., 1969; Kuriatov et al., 1984), thus allowing the use of fluorocontaining amino acids as suitable intrinsic chromophores in peptides.

The spectroscopic properties of fluorosubstituted tyrosine are the least investigated. In this work fluorescence and absorption spectra of 3-fluorotyrosine (3-F-Tyr) in aqueous solutions with various phosphate concentrations have been studied. It is known the phosphate to be important component of buffers, besides it is the specific quenching probe for analysis of the degree of exposure of tyrosine residues and peptide conformation (Shimizu et al., 1979; Ross et al., 1986).

### Material and methods

Emission spectra of 3-F-Tyr were obtained with Perkin-Elmer Model 650-10 spectrofluorometer, Absorption spectra were recorded on a Specord UV-Vis Model 40 spectrophotometer. 3-F-Tyr was synthesized and purified by the method of English (1940). Solutions under investigation were prepared by mixing appropriate aliquots of the initial solutions: 1 mM 3-F-Tyr and 3 M K<sub>2</sub>HPO<sub>4</sub>. Before measurements pH of solutions were adjusted to the required value with a pH-meter by addition of concentrated HCl (10M) or KOH (10M). All solutions contained 0.1 M KCl and were made up in twice distilled water.

## Results and discussion

3-F-Tyr fluorescence spectra are shown in Fig. 1. In the absence of phosphate the emission maximum is near 315 nm, that is a 10 nm longer wavelength shift compared with tyrosine at 305 nm (Lakowicz, 1983). Increasing phosphate concentration results in a progressive decrease in 3-F-Tyr emission at 315 nm. Concurrently, a new emission is observed at 350 nm, the emission at 315 nm being quenched practically completely at phosphate concentration 2.9 M. Similarly to tyrosine (Alev-Behmoaras et al., 1979), this new emission is a consequence of 3-F-Tyr-phosphate intraction, that is 3-fluorotyrosinate ion formation in its excited state. As it was shown earlier by Shimizu et al. (1979), Alev-Behmoaras (1964), Feitelson (1964), excited state ionization of tyrosine is facilitated by the presence of nearby proton acceptors, particularly phosphate ions. Hydrogen bonding between this acceptor and the hydroxyl moiety on 3-F-Tyr can occur in the ground state, resulting in complex poised for reaction upon excitation of 3-F-Tyr. Complex formation between 3-F-Tyr and phosphate results in red shift of absorption spectrum (about 5 nm to long waves, 2.9 M K<sub>2</sub>HPO<sub>4</sub>), leading to potential confusion with tryptophan fluorescence (Lakowicz,

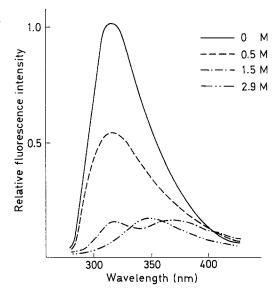


Fig. 1. Fluorescence spectra of-3-fluorotyrosine. The numbers refer to the concentration of  $K_2HPO_4$ . The pH was 7.4,  $t=20^{\circ}C$ . Excitation wavelength was 275 nm

1983). Quantitative analysis of this complexation have been carried out by the method of difference UV-spectroscopy (Moon, 1965):

$$\frac{1}{\Delta A} = \frac{1}{\Delta A_{\text{max}} K_{\text{ass}}[Q]} + \frac{1}{\Delta A_{\text{max}}},\tag{1}$$

where  $\Delta A$  – apparent optical density difference in the presence of ligand of [Q] concentration,  $\Delta A_{max}$  – maximal optical density difference with completely complexed 3-F-Tyr,  $K_{ass}$  – association constant. The plot  $1/\Delta A_{max}$  versus 1/[Q] is linear, so from eq. (1)  $K_{ass} = 0.4 \pm 0.1$  M<sup>-1</sup> is in good agreement with that of tyrosine (0.5 M<sup>-1</sup>), obtained earlier by Alev-Behmoaras (1979). Thus, 3-F-Tyr forms a complex with phosphate in the ground state, promoting the generation of 3-fluorotyrosinate ion in excited state.

The ground state pK<sub>a</sub> of 3-F-Tyr hydroxy-group (evaluated by spectroscopic titration) is near 8.3 [in peptides, according to Kuriatov et al. (1984), this value is somewhat higher, 8.9] being lower pK<sub>a</sub> than that of tyrosine [10, (Alev-Behmoaras et al. (1979)]. The first excited state pK<sub>a</sub>\* of 3-F-Tyr is 4 [calculations have been made according to Förster's cycle, Sverdlova (1985)] also being lower than the pK<sub>a</sub>\* of tyrosine 5.2–5.7, Shimizu et al. (1979). For this reason hydroxy-group deprotonation in the excited state should be (even at neutral pH) easier for 3-F-Tyr compared with tyrosine. Apparently the presence of phosphate cannot only promote this process but stabilize 3-fluorotyrosinate ion in the excited state.

Thus, quenching of 3-F-Tyr by phosphate is accompanied by a new emission peak near 350 nm due to hydroxyl moiety deprotonation. Hydrogen bonding between phosphate and 3-F-Tyr in the ground state promotes 3-fluorotyrosinate ion formation in the excited state. Selective excitation of 3-fluorotyrosinate can give emission at longer wavelengths resulting in further confusion with tryptophan fluorescence.

#### References

Alev-Behmoaras T, Toulmé J-J, Héléne C (1979) Quenching of tyrosine fluorescence by phosphate ions. A model study for protein – nucleic acid complexes. Photochem Photobiol 30: 533-539

Chen RF, Edelhoch H, Steiner RF (1969) Fluorescence of proteins. In: Leach SJ (ed) Molecular biology. Physical principles and techniques of protein chemistry, part A. Academic Press, New York London, pp 171–244

English J Jr, Mead JF, Niemann (1940) The synthesis of 3,5-difluoro- and 3-fluoro-5-iododl-tyrosine. J Am Chem Soc 62: 350–354

Feitelson J (1964) On the mechanism of fluorescence quenching. Tyrosine and similar compounds. J Chem Ptys 68: 391–397

Ghosh R, Bachofen R, Hauser H (1985) Incorporation of <sup>19</sup>F-substituted aromatic amino acids into membrane proteins from chromophores of *Rhodospirillum rubrum* 69<sup>+</sup>. FEBS Lett 188: 107–111

Kuriatov AB, Ovechkina GV, Alonycheva TN, Minajeva LP, Tsetlin VI (1984) Biosynthetic <sup>19</sup>F and <sup>14</sup>C derivatives of bacteriorodopsyn. Bioorganicheskaya Khimia 10: 333–340 (in Russian)

Lakowicz JR (1983) Principles of fluorescence spectroscopy, 1st edn. Plenum Press, New York London, p 359

- Moon AJ, Poland DC, Sherega HA (1965) Thermodinamic data from fluorescence spectra. I. System phenol-acetate. J Phys Chem 69: 2960–2966
- Ross JBA, Laws WR, Buku A, Sutherland IC, Wyssbrod HR (1986) Time-resolved fluorescence and <sup>1</sup>H NMR studies of tyrosyl residues in oxytocin and small peptides: correlation of NMR-determined conformations of tyrosyl residues and fluorescence decay kinetics. Biochemistry 25: 607–612
- Shimizu O, Watanabe J, Imakubo K (1979) Effect of phosphate ion on fluorescent characteristics of tyrosine and its conjugate base. Photochem Photobiol 29: 915–919
- Sverdlova OV (1985) Electronic spectra in organic chemistry, 1st edn. Khimia, Leningrad, p 167 (in Russian)

Authors' addresses: S. I. Vdovenko, Institute of Bioorganic and Oil Chemistry, Academy of Sciences of Ukraine, Murmanskaja street, 1 Kiev-94, 253660, Ukraine.

Received March 17, 1992